

Hydrogen Sulfide Provides Intestinal Protection During A Murine Model of Experimental Necrotizing Enterocolitis

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ABSTRACT:

Background: Necrotizing enterocolitis (NEC) continues to be a morbid surgical condition among preterm infants. Novel therapies for this condition are desperately needed. Hydrogen sulfide (H_2S) is an endogenous gasotransmitter that has been found to have beneficial properties. We therefore hypothesized that intraperitoneal injection of various H_2S donors would improve clinical outcomes, increase intestinal perfusion, and reduce intestinal injury in an experimental mouse model of necrotizing enterocolitis.

Methods: NEC was induced in five-day-old mouse C57BL/6 mouse pups through maternal separation, formula feeding, and intermittent hypoxic and hypothermic stress. The control group (n=10) remained with their mother and breastfed ad lib. Experimental groups (n=10 / group) received intraperitoneal injections of phosphate buffered saline (PBS) vehicle or one of the following H_2S donors: (1) GYY4137, 50 mg/kg daily; (2) Sodium sulfide (Na_2S), 20 mg/kg three times daily; (3) AP39, 0.16 mg/kg daily. Pups were monitored for weight gain, clinical status, and intestinal perfusion via transcutaneous Laser Doppler Imaging (LDI). After sacrifice on day nine, intestinal appearance and histology were scored and cytokines were measured in tissue homogenates of intestine, liver, and lung. Data were compared with Mann-Whitney and $p<0.05$ was considered significant.

Results: Clinical score and weight gain were significantly improved in all three H_2S -treated groups as compared to vehicle ($p<0.05$ for all groups). Intestinal perfusion of the vehicle group was 22% of baseline while the GYY4137 group was 38.7%

($p=0.0103$), Na_2S was 47.0% ($p=0.0040$), and AP39 was 43.0% ($p=0.0018$). The vehicle group had a median histology score of 2.5, while the GYY4137 group's was 1 ($p=0.0013$), Na_2S was 0.5 ($p=0.0004$), and AP39 was 0.5 ($p=0.0001$). Cytokine analysis of the intestine of the H_2S -treated groups revealed levels closer to breastfed pups as compared to vehicle ($p<0.05$ for all groups).

Conclusion: Intraperitoneal administration of H_2S protects against development of NEC by improving mesenteric perfusion, and by limiting mucosal injury and altering the tissue inflammatory response. Further experimentation is necessary to elucidate downstream mechanisms prior to clinical implementation.

INTRODUCTION:

Necrotizing enterocolitis (NEC) is the leading cause of morbidity and mortality from gastrointestinal disease in the premature neonatal population. Incidence has been estimated to vary from 12% to 20% of preterm infants, with mortality as high as 40% of those affected [1-4]. Despite NEC being the most common acquired gastrointestinal emergency among neonates, relatively little advancement has been made in available medical therapies over the last few decades. Surgical resection of necrotic bowel is required in up to 50% of cases [5]. Morbidities are significant, including intestinal stricture and short bowel syndrome as well as growth retardation and neurodevelopmental delay [3]. Therefore, a novel medical therapy for this condition is desperately needed.

One recently appreciated therapy for ischemic tissue injury has been hydrogen sulfide gas (H_2S). Recently lauded alongside nitric oxide and carbon monoxide as one of the three major gasotransmitters, it has been found to have anti-apoptotic, antioxidant, anti-inflammatory, and vasodilatory properties [6]. Though many cells produce it endogenously, it can also be applied exogenously with similar effects. In animal models, H_2S administration has been shown to provide protection against ischemic injury in the myocardium and brain [4, 7-9]. In mouse studies of intestinal ischemia and reperfusion, H_2S was applied directly to the ischemic bowel resulting in improved intestinal perfusion and decreased histologic injury secondary to the insult [4]. Additionally, Li et al has demonstrated in vitro protection in intestinal epithelial cells [10]

H_2S exists in its native form as a gas, but various substances act as H_2S donors in aqueous solution. These can be classified further into inorganic sulfide salts and

synthetic molecules. Sodium hydrosulfide (NaHS) and sodium sulfide (Na₂S) are the two most commonly used sulfide salts, and are often sold in their hydrated forms. Two frequently used synthetic molecules are GYY4137 (morpholin-4-ium 4-methoxyphenyl(morpholino) phosphinodithioate) and AP39 ((10-oxo-10-(4-(3-thioxo-3H-1,2-dithiol-5yl)phenoxy)decyl) triphenylphosphonium bromide) [11]. These are both relatively large molecules that produce hydrogen sulfide at a consistent rate over time, leading to stable concentrations in aqueous solution [12, 13]. It is thought that AP39 acts specifically on the mitochondria, while GYY4137 is more generalized [14]. All of these donors have been used in animal experiments without prohibitive adverse effects [12, 13, 15].

NEC is characterized not only by intestinal injury, but also dyscrasias in local and systemic inflammatory cytokine cascades. Interleukin 6 (IL-6), typically thought of as an acute phase reactant, has both inflammatory and anti-inflammatory properties, and may act in a protective manner against intestinal injury [16, 17]. Interleukin 1 β (IL-1 β), another acute phase reactant, is produced by many cell types, including the endothelium, in response to stress [18]. Meanwhile, a variety of angiogenic growth factors, such as vascular endothelial growth factor (VEGF), are released by the endothelium to circumvent hypoxic injury [19]. Additionally, interleukin 12 (IL-12) released by the mucosa is thought to be directly related to bacterial clearance and mucosal permeability and therefore is an important marker in experimental NEC [20]. These four signaling molecules are some of the many that are affected in experimental NEC.

Development of NEC is certainly multifactorial, but one contributor seems to be intestinal ischemia and necrosis [3]. Because of this, it is plausible that H₂S donors would be beneficial for the treatment of NEC. We therefore hypothesized that intraperitoneal injection of H₂S donors would improve clinical outcomes, increase intestinal perfusion, and reduce intestinal injury in an experimental mouse model of necrotizing enterocolitis.

MATERIALS AND METHODS

Experimental NEC Model:

Indiana University Institutional Animal Care and Use Committee approved the experimental protocol and animal use. We used a previously well published and validated model of experimental NEC [21, 22]. C57BL/6 mouse pups (bred in house from adult mice obtained from Jackson Labs, Bar Harbor, ME) were permanently separated from their mother on postnatal day (P) five. They were housed in a neonatal incubator from P5 to P9 with humidity 40% and temperature 32°C. The control group (n=10) remained with their mother and breastfeed ad libitum. Experimental groups (n=10 per group) were gavage fed with a 2 French catheter three times daily with hyperosmolar formula. Formula was prepared using 4g of Esbilac canine supplement and 6g Similac in 20 mL of nanopure filtered water (Barnstead Nanopure, APS Water Services Inc., Van Nuys, CA). Formula was discarded and prepared fresh every 48 hours. Animals were fed 300 kcal/kg/day and all feeds were supplemented with 8 mg/kg lipopolysaccharide (lipopolysaccharides from *Escherichia coli* O111:B4, Sigma-Aldrich Company LLC, Dorset, UK). Before each feed, pups were placed in a chamber

with 5% O₂ and 95% N₂ for 10 minutes. After the morning and evening feed, pups were placed in the 4°C refrigerator for 10 minutes. Mice who died less than 24 hours into the protocol were excluded, as their death was more likely due to causes other than NEC. Any animal whose tissue was liquefied or unusable for histologic evaluation (i.e. died overnight and discovered hours later) was excluded completely from evaluation. Other animals that died during the study but were identified immediately were still included in analysis for all data points.

H₂S-donor Administration:

Before hypoxic stress, experimental groups received intraperitoneal injections of phosphate buffered saline (PBS) vehicle or one of the following H₂S donors in PBS solution: (1) GYY4137, 50 mg/kg daily; (2) Na₂S, 20 mg/kg three times daily; (3) AP39, 0.160 mg/kg daily. Na₂S was dosed more frequently because as a salt in aqueous solution, its H₂S production is known to be short-lived, while the synthetic donors produce at least 24 hours of a stable H₂S concentration in aqueous solution [12, 23]. Total volume of each injection was 10 µL. Control (breastfed) groups did not receive any intraperitoneal injections. Doses are based on previous animal model studies of cardiac ischemia and reperfusion [15].

Clinical Assessment:

Control groups were assessed daily, while experimental groups were assessed with each feed. Pups were assessed in a systematic fashion to ensure consistency.

The clinical sickness score was previously published by Zani [21]. The reported clinical assessment score was the pup's last score prior to death or euthanasia (Table 1).

Perfusion Analysis:

Intestinal perfusion was analyzed transcutaneously with a Laser Doppler Perfusion Imager (LDI; Moor Instruments, Wilmington, DE). As the laser was known to penetrate skin up to several millimeters thick, our previously published protocols were modified to allow for transcutaneous acquisition in the neonatal pups in order to avoid multiple surgeries [4]. Pups were held supine by their front limbs and tail and images were acquired of the entire torso and abdomen. Using the computer software, a region of interest was created from the diaphragm to the pelvis to obtain a flux mean perfusion value within this region. Full, visible bladders were excluded from the region of interest for consistency. Perfusion was expressed as a percentage of baseline perfusion on P5. Pups that died during the protocol (1 in vehicle group, 1 in Na₂S group, and 1 in AP39 group) were included in evaluation with a perfusion value of 0%.

Macroscopic Gut Assessment and Histology:

On P9, pups were sacrificed by decapitation and intestinal appearance was assessed with a macroscopic scoring system based on color, consistency, and dilation of the intestine as described by Zani et. al. (Table 2). The terminal ileum and distal jejunum was then formalin fixed, paraffin embedded, and stained with hematoxylin and eosin. Two blinded authors evaluated degree of injury by histology. The histological scoring system used was published by Zani et. al [21] with scores ranging from 0 to 4.

0 = normal intestine; 1 = some disarrangement of villus enterocytes, villus-core separation; 2 = significant disarrangement of villus enterocytes, villus-core separation down sides of villi, blunting of villi; 3 = epithelial sloughing of villi, loss of villi; 4 = necrosis. A score of 2 or higher indicated presence of NEC, with 3 or higher indicating severe NEC.

Intestinal Cytokine Analysis:

Murine organs were acquired and processed for proteins as previously described [4, 24, 25]. Following euthanasia, part of the small intestine, liver, and lung was snap frozen in liquid nitrogen and stored at -80°C. Tissue was thawed and homogenized with the Bullet Blender (Next Advance, Averill Park, NY) in RIPA buffer (Sigma, St. Louis, MO) with 1:100 dilutions of both phosphatase and protease inhibitors (Sigma, St. Louis, MO). After homogenization, samples were centrifuged at 12,000 rpm and supernatants were collected for further analysis. Total protein was quantified with the Bradford Assay using a spectrophotometer (VersaMax microplate reader, Molecular Devices, Sunnyvale, CA).

Murine interleukin 6 (IL-6), vascular endothelial growth factor (VEGF), interleukin 1 β (IL-1 β), and interleukin 12 (IL-12) were measured using ELISA (R&D Systems, Bio-Techne Corporation, Minneapolis, MN). ELISAs were repeated to ensure consistency and values were normalized to the control group. Assays were performed at 1:20 dilution and cytokine concentrations were normalized to total protein for each sample.

Statistical Analysis:

Ordinal data was reported using median and interquartile range. Continuous variables were reported as mean \pm SEM. All non-parametric data was compared using the Mann-Whitney U test. GraphPad Prism 7 (GraphPad Software, La Jolla, CA) was used for all statistical analysis and figures. P values less than 0.05 were considered statistically significant.

RESULTS

H₂S improves weight gain and clinical status in experimental NEC.

The breastfed group gained an average of 2.23 ± 0.10 grams over 4 days, while the PBS-treated NEC group averaged a gain of 0.01 ± 0.02 grams ($p < 0.0001$).

Although the overall weight gain was minimal, compared to the vehicle treated NEC group, each H₂S-treated NEC group gained significantly more weight. The GYY4137 group gained 0.16 ± 0.03 grams ($p = 0.0007$), the Na₂S group gained 0.15 ± 0.05 grams ($p = 0.0142$), and the AP39 group gained 0.09 ± 0.02 grams ($p = 0.0146$, Figure 1A)

Clinical assessment scores were significantly worse in the vehicle NEC group compared to the breastfed animals and improved in the H₂S-treated groups (Figure 1B). While the median score in the control group was 0 (IQR=0-0), that of the vehicle NEC group was 3 (IQR=1.75-5.25, $p = 0.0001$). The median score for the H₂S-treated NEC groups were lower: GYY4137: 1 (IQR=0-2, $p = 0.0109$), Na₂S: 1 (IQR=0-1, $p = 0.0027$), and AP39: 1 (IQR=1-2, $p = 0.0185$).

H₂S improves intestinal perfusion in experimental NEC.

At P9, the mean perfusion of the breastfed group was $54.35\% \pm 5.28\%$ of baseline on P5. The vehicle group's P9 perfusion was $24.16\% \pm 4.45\%$ which was significantly lower than the control group ($p=0.0015$). The H_2S -treated NEC groups had higher perfusion than the vehicle NEC group (Figure 2). At P9, perfusion of the GYY4137 group was $38.69\% \pm 2.03\%$ ($p=0.0079$), Na_2S was $46.95\% \pm 7.93\%$ ($p=0.0162$), and the AP39 group was $37.48\% \pm 5.24\%$ ($p=0.0296$).

H_2S reduces intestinal injury in experimental NEC.

Median macroscopic injury score of the breastfed group was 0 (IQR=0-0) and that of the vehicle treated NEC group was 2.5 (IQR=2-3, $p<0.0001$). The median score of the GYY4137 group was 1 (IQR=0-1, $p<0.0001$), the Na_2S group was 0.5 (IQR=0-1, $p<0.0001$) and the AP39 group was 1 (IQR=0-2.25, $p=0.0103$, Figure 3A). Histologic injury followed a similar pattern with a median score of 0.5 (IQR=0-1) for the breastfed pups and 2.5 (IQR=1.5-3) for vehicle-treated NEC pups ($p<0.0001$). All 3 H_2S -treated groups had improved histology compared to vehicle. The GYY4137 group's median score was 1 (IQR=1-1.625, $p=0.0011$), the Na_2S group's was 0.5 (IQR=0.5-0.125, $p=0.0004$), and the AP39 group's was 0.5 (IQR=0.5-1.125, $p=0.0001$). Incidence of NEC in the Vehicle group was 70%, and it was 20%, 10%, and 0% in the GYY4137, Na_2S , and AP39 groups respectively. Severe NEC was present in 30% of the vehicle-treated animals, and 0% of the H_2S -treated animals (Figure 3B).

H_2S improves local inflammation in the intestine during experimental NEC.

Intestinal cytokines and growth factors were altered in the experimental NEC model. Animals in the vehicle group had significant derangements in intestinal IL-6, VEGF, IL-1 β and IL-12 compared to breastfed control (Figure 4). IL-6, IL-1 β and IL-12 were all significantly decreased, while VEGF was increased in the vehicle treated NEC group. Treatment with each of the H₂S donors brought these cytokines back toward control levels.

H₂S improves systemic inflammation during experimental NEC.

Liver and lung cytokines and growth factors were also affected in the experimental NEC model with significant deviations in the vehicle group as compared to control (Figure 5). In both lung and liver, IL-6, VEGF, and IL-1 β were significantly reduced in the vehicle treated NEC group compared to control. Only treatment with GYY4137 resulted in statistically significant improvements in IL-6 and VEGF in both the lung and liver.

DISCUSSION

Necrotizing enterocolitis is a devastating condition of the premature neonate that often requires extensive surgical resection of affected intestine for adequate treatment. Consequences of this disease result in high morbidity and mortality rates across this population. A medical therapy that would reduce the need for surgical treatment is certainly needed. Herein, we demonstrated that treatment with hydrogen sulfide (GYY4137, Na₂S, and AP39) can provide protection in a mouse model of experimental

NEC by improving mesenteric perfusion, reducing mucosal injury, and by normalizing local and systemic cytokine and growth factor production.

H₂S has recently been appreciated as a gas mediator like carbon monoxide and nitric oxide. Though it is synthesized endogenously from the amino acid cysteine, it can also be administered exogenously via an H₂S donor. H₂S donor administration has been shown to be beneficial in experimental models of ischemic injury [4, 6, 15, 26, 27]. Because NEC is mediated, at least in part, through intestinal ischemia and necrosis, H₂S therapy may be of benefit in this situation as well.

We noted an improvement in overall clinical status, including weight gain, in mice treated with H₂S. It is likely that these animals are not only healthier due to decreased systemic inflammation, but also may absorb nutrients more efficiently due to less diseased bowel. It has previously been postulated that local and systemic inflammation during NEC is at least partly responsible for the known sequelae of weight loss and clinical deterioration in neonates [28].

Improvement in intestinal perfusion was another observed benefit of H₂S donor administration. H₂S is thought to modulate intestinal blood flow, in part as a direct intracellular messenger, but additionally through nitric oxide (NO) mediated effects [4]. Both H₂S and NO are potent smooth muscle relaxants, likely leading to vessel dilatation and improved mesenteric perfusion [29]. One of the possible mechanisms for this interaction involves H₂S mediated upregulation of endothelial nitric oxide synthase (eNOS). H₂S has been shown to sulfhydrylate critical cysteine residues on eNOS, thereby causing enzyme dimerization and upregulation of NO production [30]. Other

studies have demonstrated that eNOS is an integral intermediary in hydrogen sulfide's ability to improve intestinal perfusion [4].

Bowel injury was also reduced both macroscopically and histologically compared to vehicle control. While this is surely due in part to improved mesenteric perfusion, other beneficial actions of H₂S may be at play as well. Other targets of protein sulfhydrylation and modification include potassium channels, phosphatases, transcription factors, and other integral cell-signaling pathways [31, 32]. Sulfhydrylation can also directly protect proteins and preserve function by preventing oxidation at cysteine residues [33]. Additionally H₂S has anti-apoptotic and anti-inflammatory properties, mediated through the NF- κ B pathways [34]. These combined effects may have a direct effect on the intestinal epithelium outside of its effects on the mesenteric vasculature.

The experimental NEC model also led to significant derangements in cytokines and growth factors in the intestine. IL-6 was decreased in our NEC model, but returned to near breastfed control levels after the application of H₂S donors. Though IL-6 is usually thought of as pro-inflammatory, some studies have suggested that it plays a more protective role in the intestine and acts as an anti-inflammatory mediator. This effect is likely mediated through its influence on other cytokines such as IL-1 [35]. One additional possibility is that by the time the tissue is harvested, the IL-6 has been exhausted from severe inflammation over multiple days. Further investigation into IL-6's role in intestinal ischemic injury is needed.

VEGF is produced by endothelial cells in response to hypoxia or stress [18]. It was elevated in the vehicle group, but reduced by treatment with H₂S donors. This

would suggest that the endothelium was less stressed after administration of hydrogen sulfide donors. VEGF is also considered a growth factor and it plays a role in regulation of angiogenesis [19]. The results of this study might suggest that a higher level of angiogenesis was not necessary following H₂S administration.

IL-1 β was consistently decreased in our NEC model, and returned to near normal levels with H₂S treatment. IL-1 β is a secretory molecule that is released from macrophages. It is typically pro-inflammatory, however, has not been shown to be elevated in NEC in previous studies [36]. It also plays a role in gut integrity and enteric mucosal protection by stimulating glial cells [37]. It therefore stands to reason that it may be reduced in experimental NEC, when the intestine is injured and gut barrier permeability is higher than baseline.

IL-12 was decreased in the vehicle group, and increased toward baseline with H₂S administration. IL-12 helps to promote the actions of interferon gamma and has been implicated as a factor in the onset of NEC and the progression toward fulminant disease [16]. Studies have demonstrated decreased IL-12 production in the intestinal epithelium in mice during experimental NEC [20]. This is consistent with our results, and improvement secondary to H₂S administration is a positive indicator that the pathology was resolving.

In the lung and liver tissue, similar patterns were noted in the vehicle treated NEC group compared to control with regard to IL-6 and IL-1 β . However, VEGF decreased in the vehicle group compared to control. IL-12 was not evaluated in the liver and lung because it is thought to be more of an intestinal marker for NEC [20]. Treatment with GYY4137 did result in an improvement in IL-6 and VEGF, but the rest of

the donors did not appear to facilitate a significant improvement in the systemic inflammatory response. These data indicate that despite the improvement in intestinal injury that resulted from direct application to threatened bowel, the systemic effects of experimental NEC were likely not completely ameliorated by this therapy. Further investigation into the effects of H₂S on systemic inflammation is therefore warranted.

LIMITATIONS

Since the exact cause of NEC is not completely understood, we are limited in the types of animal models that are available for use. As such, clinical correlation to human illness may be limited. Although the clinical scores and intestinal perfusion parameters are strong indicators of the development of NEC, the diagnosis can only be established at postmortem evaluation of the intestine, and this makes it difficult to assess during the experiment whether or not NEC is being prevented. Additionally, it is difficult to blind clinical assessments of the animals, as they were performed by the same person who administered the medication. This was remedied by a very structured approach to clinical assessment, in addition to including more objective measures like intestinal perfusion and cytokine analysis. All macroscopic and histologic evaluation was blinded, and corresponded with other clinical assessments.

CONCLUSION

H₂S administration is beneficial in experimental NEC. Mouse pups treated with H₂S experienced only marginal weight gain, but did exhibit improved clinical status, increased intestinal perfusion, reduced intestinal injury, and normalization of inflammatory markers as compared to vehicle. Further experimentation into

mechanism, mode of delivery, and adverse effects is necessary prior to widespread therapeutic use.

FIGURE LEGENDS

Figure 1: *H₂S improves weight gain and clinical status in experimental NEC.* A) The vehicle group had virtually no weight gain compared to control, and though the H₂S groups had very minimal, it was statistically better than the vehicle group. B) Clinical assessment scores were higher (worse) in the vehicle group compared to control, and improved in the H₂S-treated groups. Transverse bars represent median clinical score and each symbol represents an individual pup's score. (*: p<0.05 vs. breastfed, #: p<0.05 vs. vehicle)

Figure 2: *H₂S improves intestinal perfusion in NEC as measured by LDI compared to vehicle.* A) Perfusion on P9 is reported as a percentage of baseline perfusion on P5. Perfusion was significantly lower in the vehicle group and improved in all three H₂S-treated groups. (*: p<0.05 vs. breastfed, #: p<0.05 vs. vehicle) B) Representative pictures from LDI software of each group's transcutaneous intestinal perfusion. Red areas indicated higher perfusion, and blue areas indicated lower levels of perfusion. The left side of each image is the pelvis and the right is the diaphragm.

Figure 3: *H₂S reduces intestinal injury in NEC compared to vehicle.* A) Macroscopic injury score was higher (worse) in the vehicle group as compared to the breastfed animals. All three H₂S-treated groups had improved scores. B) Histologic injury scores were higher (worse) in the vehicle group compared to the breastfed group. All three

H₂S-treated groups had improved scores. (*: p<0.05 vs. breastfed, #: p<0.05 vs. vehicle) C) Representative sections from each treatment group (H&E stain, 20x).

Figure 4: *H₂S normalizes intestinal inflammation in NEC compared to vehicle.* The vehicle group had significantly decreased IL-6, IL-1 β , and IL-12 and increased VEGF compared to control. Each H₂S-treated group had levels closer to breastfed control indicating normalization in local inflammation. (*: p<0.05 vs. breastfed, #: p<0.05 vs. vehicle)

Figure 5: *H₂S normalizes systemic inflammatory response in NEC compared to vehicle.* The vehicle group had significantly decreased IL-6, VEGF, and IL-1 β compared to control. H₂S treatment may have facilitated some normalization back to breastfed control levels, however this was not a statistically significant difference, with the exception of the GYY4137 group for IL-6 and VEGF. (*: p<0.05 vs. breastfed, #: p<0.05 vs. vehicle)

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Figure 1

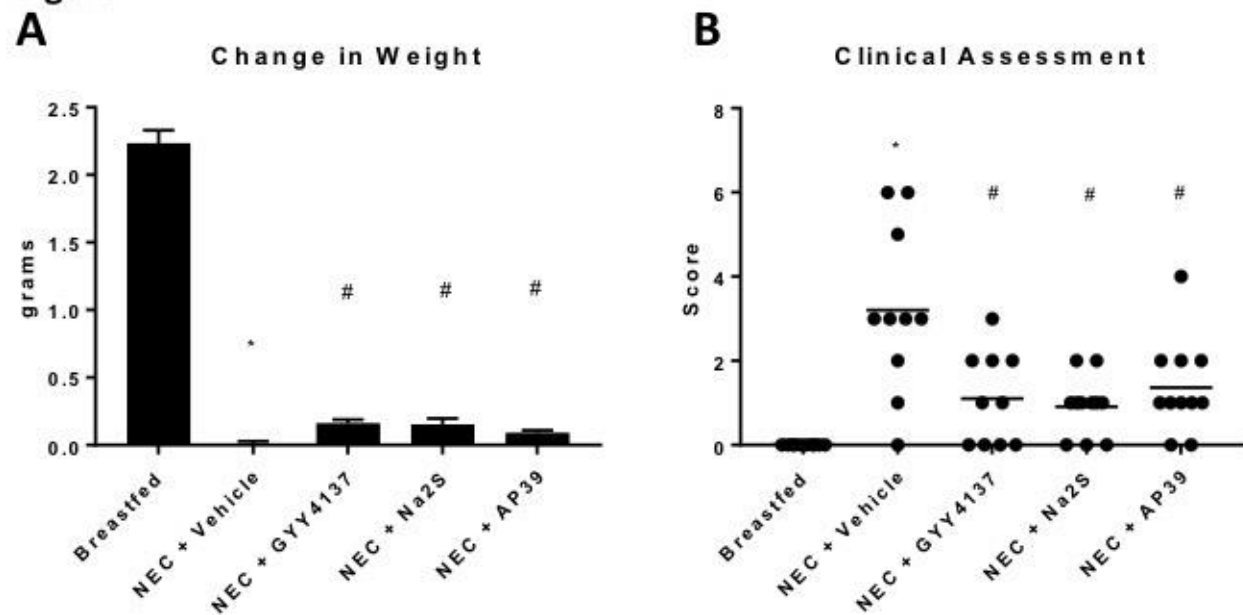
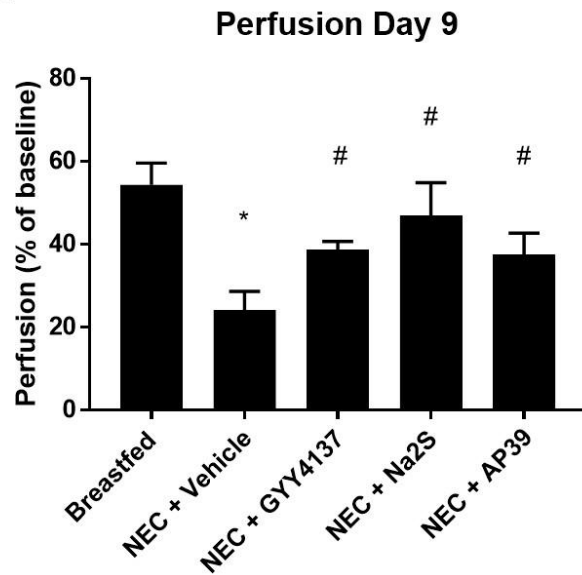


Figure 2

A



B

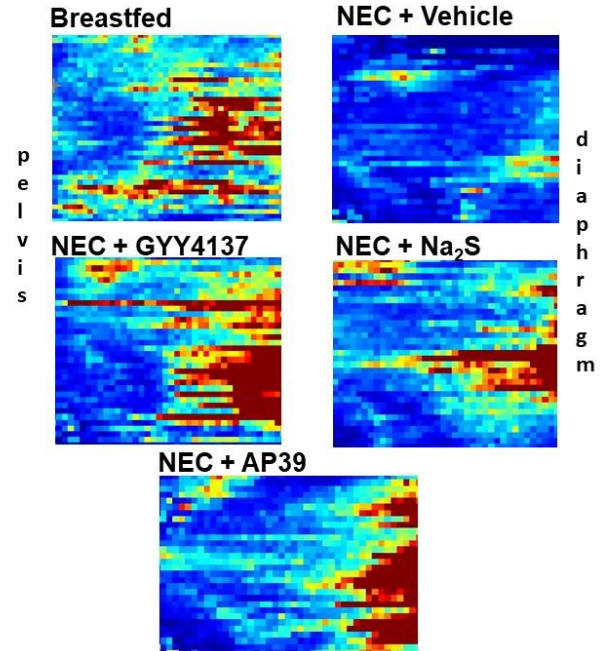


Figure 3

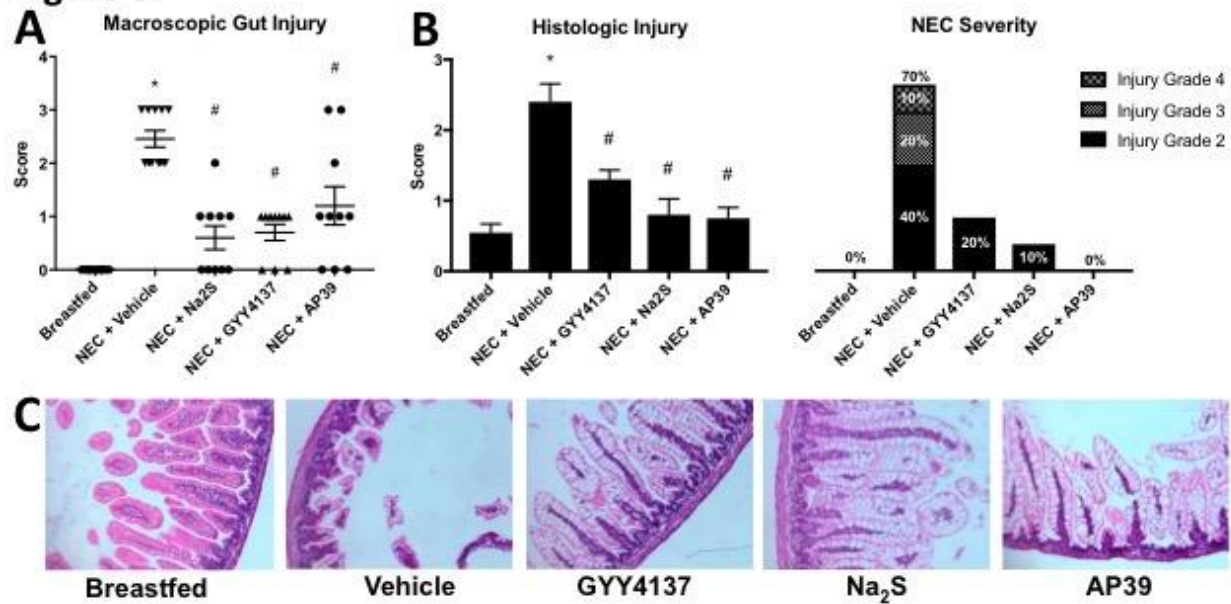


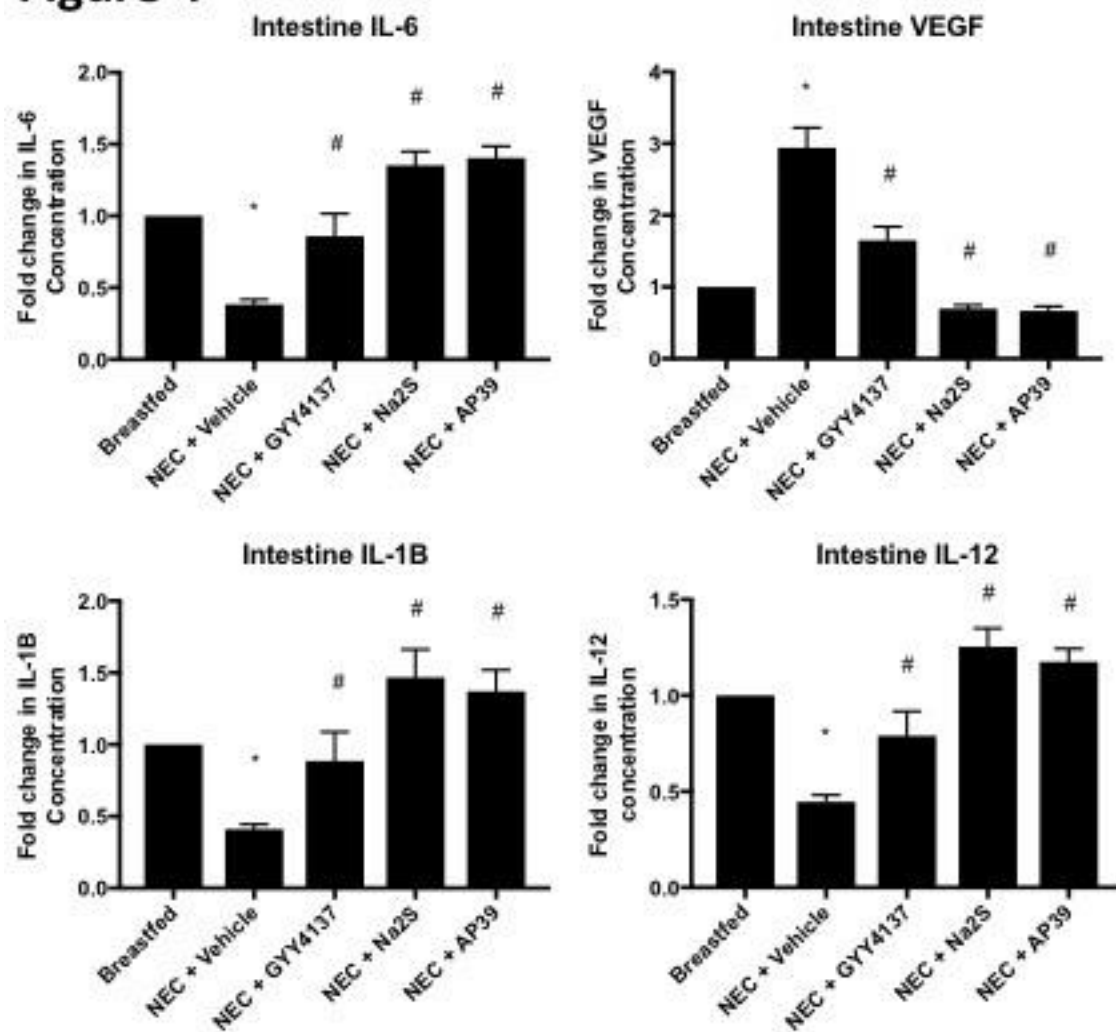
Figure 4

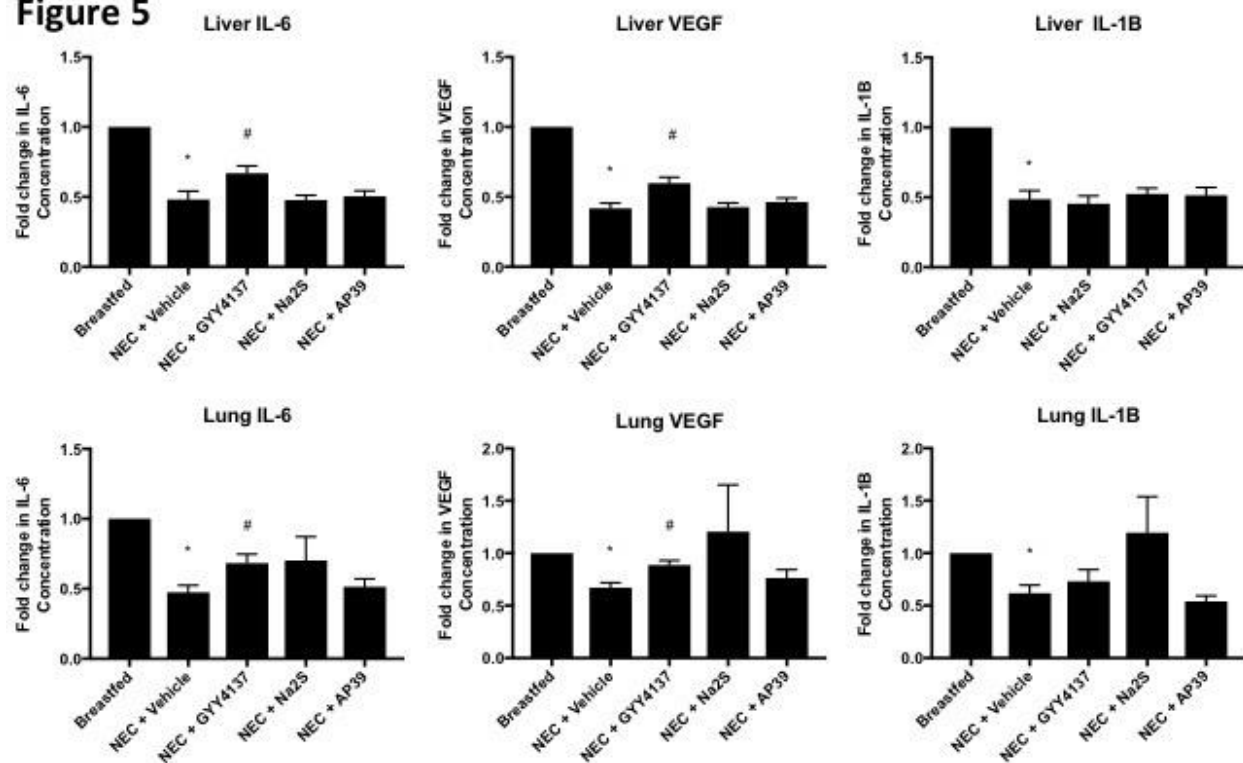
Figure 5

Table 1. Clinical Assessment Score	
Appearance	
0	Tonic and well-hydrated, glossy skin
1	Slimmer, with visible skin folds
2	Skinny, dehydrated
3	Gasping, in agony
Natural Activity	
0	Moving normally in the cage
1	Wiggles if put supine
2	Does not wriggle if put supine
3	Not moving limbs, lying still, supine or lateral decubitus
Response to Touch	
0	Alert without any stimulation
1	Responding to mild stimulation (gentle foot touch)
2	Responding to vigorous stimulation (foot pinch)
3	Unresponsive to vigorous stimulation (does not become alert when picked up and moved)
Body Color	
0	Pink
1	Pale or dark distal extremities, fingers/toes
2	Pale whole body
3	Grey

Zani et. al. 2008

Table 2. Macroscopic Gut Assessment Score	
Consistency	
0	Normal, does not tear when dissecting from surrounding tissue
1	Moderately friable
2	Liquefied, jelly-like, extremely friable
Color	
0	Normal, yellow/tan
1	Patchy discoloration, pink, congested
2	Extensive discoloration, dark, necrotic
Dilation	
0	Normal, no dilation
1	Patchy areas of dilation
2	Extensive dilation throughout small bowel

Zani et. al. 2008